Fosfomycin Sensitivity Pattern among Uropathogens Isolated from Patients Visiting Day Care Facility of Sushila Tiwari Hospital in Kumaun Region, Uttarakhand, India

**Microbiology Section** 

SHRADDHA SHARMA<sup>1</sup>, PANKAJ KUMAR VERMA<sup>2</sup>, VINITA RAWAT<sup>3</sup>, UMESH<sup>4</sup>, VIKRANT NEGI<sup>5</sup>

# (CC) BY-NC-ND

# ABSTRACT

**Introduction:** Urinary Tract Infections (UTI) are one of the most common infections responsible for antibiotic resistance. There are limited antibiotics options for treating the cases due to Multi Drug Resistant (MDR) bacteria. Fosfomycin is being used for treating UTIs and has shown promising results even against MDR pathogens.

**Aim:** To determine the fosfomycin sensitivity pattern along with Minimum Inhibitory Concentration (MIC) against uropathogens by agar dilution method.

**Materials and Methods:** The present cross-sectional study was conducted in the Department of Microbiology, Government Medical College Haldwani, Uttarakhand, India, between August 2017 to September 2019. Clean catch, mid stream urine samples were inoculated on Cystine Lactose Electrolyte Deficient (CLED) agar and incubated. The significant growths of pathogenic bacteria were subjected to antibiotic susceptibility testing. Fosfomycin (200 µg)

disc was used in Kirby-Bauer disc diffusion testing. Fosfomycin trometamol MIC was determined by agar dilution method as per Clinical and Laboratory Standards Institute (CLSI) guidelines. The data collected in the study were analysed by using Statistical Package for the Social Science (SPSS) software version 20.0.

**Results:** Significant growth of pathogenic bacteria was observed in 365 out of total 2725 urine samples. *E.coli* (72.32%) was the leading isolate followed by *Enterococcus* species (10.41%). Fosfomycin was recorded as the most active antibiotic against all the bacterial pathogen with 85-100% susceptibility except *Proteus* species (40%) in disc diffusion method. The MIC of fosfomycin was recorded between 4-64 against most of the isolates by agar dilution method.

**Conclusion:** Fosfomycin is the most active antibiotic against all the uropathogens in the study setup and can be included in empirical treatment of day care patients along with nitrofurantoin.

Keywords: Agar dilution, Minimal inhibitory concentration, Multi drug resistance, Urinary tract infections

## INTRODUCTION

The UTIs are among the most commonly occurring human infections [1,2]. Approximately, 50% of women will experience UTI atleast once during their lifetime while about 25% of women will suffer recurrent infection [3]. UTIs being one of the most common human infections are the reason for large proportion of antibiotic consumption and thus contributing antibiotic resistance [4]. An increasing proportion of UTIs are due to MDR pathogens for which there are limited treatment options [5].

Reconsideration of 'neglected' antibacterial drugs is one of the approaches for facing this complicated burden of disease as older drugs like temocillin, mecillinam, fusidic acid, polymyxins etc., have documented as potentially useful against MDR pathogens [6,7]. One such agent, fosfomycin, is being called back into play in the United Kingdom (UK) for treating UTI [8,9].

Fosfomycin trometamol is a well-tolerated drug as well as have a broad spectrum of activity against a wide range of Gram-positive and Gram-negative bacteria [10]. It has minimal toxicity, and acts as a time-dependent inhibitor of the MurA enzyme, which catalyses the first committed step of peptidoglycan synthesis involving phosphoenolpyruvate synthetase. As there are no data available on the susceptibility pattern as well as MIC of fosfomycin from this part of country. Hence, the present study was conducted to assess the fosfomycin susceptibility pattern along with MIC against uropathogens by agar dilution method.

## **MATERIALS AND METHODS**

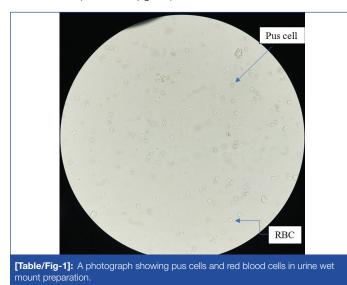
The present cross-sectional study was conducted in the Department of Microbiology, Government Medical College Haldwani, Uttarakhand,

India, between August 2017 to September 2019. Written permission was obtained from Institutional Human Ethical Committee (Letter No. 394/GMC/IEC/2017/Reg. No. 363/IEC/R-16-09-2017). Written informed consent form were signed and collected from the volunteer patients.

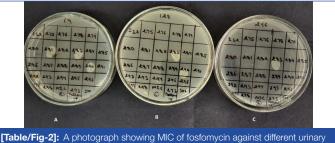
Patients visiting the Out-Patient Department (OPDs) with suspected UTIs (symptoms such dysuria with irritating voiding, urinary urgency, frequency, nocturia, painful voiding, pain after voiding, sensation of bladder fullness or lower abdominal discomfort and sometime pain in the suprapubic area) and ≥16 years of age disregarding their gender, were enrolled in the study. All the younger patients <16 years of age and the hospital admitted patient were excluded from the study. Consecutive, non-duplicate, midstream clean catch urine samples were collected in a sterile urine container from the enrolled patients with sign and symptoms along with clinical diagnosis of UTI.

Samples were transported to Microbiology laboratory and processed without any delay. In case of delay, samples were kept at 4-8°C. Wet mount preparation was made directly from the samples and observed under light microscope [Table/Fig-1]. All urine samples were plated semi-quantitatively on CLED agar and incubated at 37°C for overnight. Any suggestive growth was further tested for Gram's staining and biochemical identifications as per standard operating procedures of the laboratory. They were further subjected to antibiotic susceptibility testing by disc diffusion method and interpreted as per CLSI guidelines, 2017 [11]. The results of the standard single-disc susceptibility tests with disks containing 200  $\mu$ g of fosfomycin and 50  $\mu$ g of glucose-6-phosphate were interpreted according to CLSI 2017 guidelines. The zone size  $\geq$ 16 mm was reported Susceptible (S), while 13-15 mm as Intermediate (I) and  $\leq$ 12 mm as Resistant (R).

Agar dilution method: The isolates were subjected to MIC testing against fosfomycin trometamol by agar dilution method on Muller Hinton Agar (MHA) supplemented with 25 µg/mL of glucose-6-phosphate to reduce the rate of false resistance as per CLSI guidelines 2017 [11]. Fosfomycin trometamol was used as fosirol powder (Cipla Ltd.,). Muller Hinton Agar with different concentrations of fosfomycin (2, 4, 8, 16, 32, 64, 128, 256 µg/mL) was used. After adjusting the turbidity with 0.5 McFarland standards, 10 µL of bacterial culture of test organism was spot inoculated on MHA plate with different concentrations of fosfomycin. Plates were incubated overnight at 37°C and examined for growth. The MIC values obtained were interpreted according to the following criteria- Susceptible (S) ≤64 µg/mL, Intermediate (I)-128 µg/mL, Resistant (R) ≥256 µg/mL and *E. coli* ATCC 25922 and *E. faecalis* ATCC 51299 was used as a control strain (MIC 0.5-2 µg/mL).



Isolates not growing in A (first agar plate) have MIC  $\leq$ 64 µg/mL, so interpreted as Susceptible. Isolates growing in A but not growing on B (281 and 292) have MIC  $\leq$ 128 µg/mL, interpreted as intermediate. Isolates growing on B and C are interpreted as resistant having MIC ( $\geq$ 256 µg/mL) [Table/Fig-2].



[hable/rig-2]: A photograph showing MiC of rostomycin against different unnary isolates by agar dilution method. A=Agar containing fosfomycin trometamol concentration of 64 µg/mL.

B=Agar containing fosfomycin trometamol concentration of 128 µg/mL

C=Agar containing fosfomycin trometamol concentration of 256 µg/mL

# STATISTICAL ANALYSIS

Fisher's-exact test was performed for statistical analysis of the data obtained in the study by using SPSS version 20.0 (IBM SPSS statistics 20). A p-value <0.05 was considered statistically significant.

#### RESULTS

A total of 2,725 urine samples were tested, out of which 365 had significant growth of urinary bacterial pathogens. The male:female ratio was recorded 04:1 [Table/Fig-3]. Among 365 isolates *E. coli* dominated the list with 72.32% followed by *Enterococcus* species (10.41%) [Table/Fig-4].

The antibiotic susceptibility pattern by disc diffusion showed high resistance against cefazolin and trimethoprim/sulphamethoxazole

	No. of patients						
Age range (years)	Male	Female	Total				
16-30	31	103	134				
31-45	24	65	89				
46-60	24	54	78				
>60	40	24	64				

[Table/Fig-3]: Age and gender distribution of patients.

Organisms	Total No. (n=365)	Percentage					
E. coli	264	72.32%					
Enterococcus spp.	38	10.41%					
Klebsiella spp.	29	7.95%					
Proteus spp.	11	3.01%					
Citrobacter spp.	07	1.92%					
Enterobacter spp.	07	1.92%					
Pseudomonas spp.	07	1.92%					
Acinetobacter spp.	02	0.55%					
[Table/Fig-4]: Organism isolated from UTI patients							

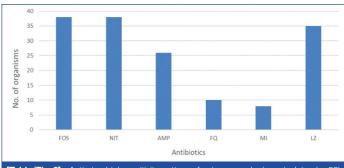
(COT) by all the pathogens except *Klebsiella* spp. and *Citrobacter* spp. The resistance against fluroquinolones was also seen in most pathogens except *Pseudomonas* spp, *Enterobacter* spp and *Proteus* spp, *E. coli* also revealed high rate of resistance towards fluoroquinolones in comparison to other pathogens. Nitrofurantoin was observed very active against members of Enterobacteriaceae

family except *Proteus* spp., which is inherently resistance. Non fermenters also revealed resistance to nitrofurantoin. Fosfomycin was recorded as most sensitive antibiotic against all the pathogens except some species of *Proteus* [Table/Fig-5].

	FOS		NIT		СОТ		CZ		FQ	
Organisms (n=327)	S	R	S	R	S	R	S	R	S	R
<i>E. coli</i> (n=264)	260	04	250	14	90	174	77	187	76	188
Klebsiella spp. (n=29)	26	03	20	09	16	13	15	14	15	14
Proteus spp. (n=11)	05	06	*NA	*NA	02	09	02	09	07	04
Citrobacter spp. (n=07)	07	00	06	01	05	02	04	03	04	03
Enterobacter spp. (n=07)	07	00	05	02	03	04	01	06	05	02
Pseudomonas spp. (n=07)	06	01	*NA	*NA	02	05	02	05	05	02
Acinetobacter spp. (n=02)	02	00	00	02	02	00	00	02	01	01
Total	313	14	281	28	120	207	101	226	113	214
p-value	<0.001		<0.001		<0.001		<0.001		<0.001	
[Table/Fig-5]: Organism-wise antibiotic sensitivity pattern by Kirby-Bauer disc-diffusion										

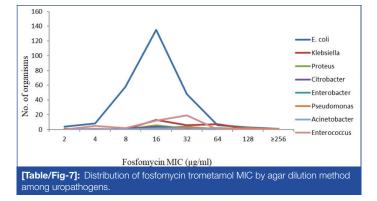
method among gram-negative uropathogens (n=327). FOS: Fosfomycin; NIT: Nitrofurantoin; COT: Trimethoprim/Sulphamethoxazole; CZ: Cefazolin; FQ: Fluoroquinolones. S: Sensitive, R: Resistant \*NA: Not applied The Fisher-Exact test was performed for comparing proportions

High rates of resistance were seen in *Enterococcus* spp. against Minocycline and Fluoroquinolones. Fosfomycin and Nitrofurantoin were 100% susceptible to *Enterococcus* spp. Linezolid and ampicillin also recorded as invitro active antibiotics against Enterococcal isolates [Table/Fig-6].



[Table/Fig-6]: Antimicrobial sensitivity pattern of enterococcal urinary isolates (n=38) FOS: Fosfomycin; NIT: Nitrofurantoin; AMP: Ampicillin; FQ: Fluoroquinolones; MI: Minocycline; L7: Linezolid

The MIC value of fosfomycin against most of the susceptible uropathogens was noted between 4-64 µg/mL [Table/Fig-7].



# DISCUSSION

The UTIs are one of the most common bacterial infections and second most common infectious disease in community and hospitals. UTIs are the most prevailing ailment affecting almost all age groups and both genders. In the present study female were found to be predominant over males among UTI patients except in geriatrics age group (i.e., >60 years). Among the 365 urinary isolates of the current study, majority were gram negative bacilli (89.58%) mainly consisting members of Enterobacteriaceae (97.24%). In the present study *E. coli* was found to be the predominant pathogen. The result was in concordance with other studies where *E. coli* were accounted as 70-80% of total urinary isolates [12,13]. There are multiple factors for *E. coli* being the most common among uropathogens such as it being as most common enteric flora and virulence factors adhesins operative through type-I fimbriae and P fimbriae which helps it to gain entry into urethra [14].

*Enterococcus* spp. was reported as second most common urinary pathogens in the present study similarly reported by few other studies [15-17]. *Enterococcus* spp. has been reported as important urinary pathogen in patients with urinary tract abnormalities and related complications [18,19]. The prevalence of *Klebsiella* spp. in the present study was (7.95%). Similar percentage of *Klebsiella* spp. were reported by others [1,17,18,20,21]. The prevalence of nonfermenters was recorded (2.75%). Similar results have been reported by few authors previously [1,22].

The treatment of UTIs varies according to the age of the patient, sex, underlying disease, infecting agent and whether there is lower or upper urinary tract involvement. Antibiotics used in the therapy of UTI are usually able to reach high urinary concentrations, which are likely to be clinically effective [23].

As per the Infectious Diseases Society of America (IDSA) guidelines cotrimoxazole is the recommended drug for the treatment of UTIs in settings where the prevalence of resistance is <10-20% and ciprofloxacin is recommended where this resistance is >20%. Co-trimoxazole was preferred as initial first line drug for treatment of UTI [24]. Moreover, *Enterococcus* species which is second most common gram-positive bacteria causing UTI is inherently resistant to co-trimoxazole [11].

The other agents used in the treatment of UTI include fluoroquinolones, cephalosporins, other  $\beta$ -lactams with or without  $\beta$ -lactamase inhibitors and nitrofurantoin. Recently, several studies have revealed increasing trends of resistance to many antimicrobials including the fluoroquinolones [25-28].

In the present study, 66% of *E. coli* was reported to be resistant towards Trimethoprim/sulfamethoxazole. Highest resistance was recorded against *Proteus spp.* (81.82%) and *Pseudomonas* spp. (71.43%). *Acinetobacter* spp. was noted as the most sensitive pathogen towards cotrimoxazole. Higher resistance rates were also reported by other authors [25,26]. However, compared to mentioned studies, Sotto A et al., have found low level (26.9%) of resistance to Trimethoprim/sulfamethoxazole [29]. The low cost, widespread availability and usage have led to increase in the resistance of

Gram-negative bacilli and also have a disadvantage as *Enterococcus* spp are inherently resistance Trimethoprim/sulfamethoxazole [11].

In the present study, (71.22%) of all *E. coli* were found to be resistant to fluoroquinolones. Ciprofloxacin resistance was comparatively less among Gram-negative uropathogens like *Pseudomonas* spp., *Acinetobacter* spp., *Enterobacter* spp. Similar results were observed by Mandal J et al. and Jain P et al., [27,28]. Fluoroquinolones resistance rate was observed 73% in *Enterococcus* spp. Similar resistance pattern was reported by Mandal J et al., in their study [27].

In the present study, resistance to nitrofurantoin has been observed in *E. coli*, and *Enterococcus* spp. i.e., 5.31% and 0%, respectively. Similar resistance pattern has been observed by Manjunath GN et al., and Keepers TR et al., [16,30]. In the present study 31.04%, *Klebsiella* spp. was found resistant to nitrofurantoin. None of the *Acinetobacter* spp. was found to be susceptible towards nitrofurantoin. Indeed, despite of its use for long time, nitrofurantoin has retained its broad-spectrum activity against gram-positive and gram-negative bacteria for UTI prophylaxis [21].

In the present study, fosfomycin showed 100% sensitivity in case of *Enterococcus* spp., *Acinetobacter* spp., *Citrobacter* spp. and *Enterobacter* spp. and resistance in *E. coli*, *Klebsiella* spp., *Pseudomonas* spp were 1.52 %, 10.35% and 14.29%, respectively. There was higher resistance seen in *Proteus* spp. 54.55% among Enterobacteriaceae family. As compared to present study, Jadoon SA et al., found higher resistance in case of *E. coli* (5%) and higher susceptibility against *Klebsiella* spp (100%) [31]. Barry AL and Fuchs PC, have reported 10% of *P. aeruginosa* strains resistant to fosfomycin while Lu CL et al., have demonstrated higher rates of resistance to fosfomycin among *P. aeruginosa* isolates [32,33]. The fosfomycin resistance in *P. aeruginosa* may occur due to over-expression of *fosA* gene by enzymatic modification of the antibiotic [34].

Out of four, *E. coli* strains which were interpreted as resistant by disc diffusion method, only one strain was found to have a value of MIC more than 256 µg/mL. A MIC value of 132 µg/mL was observed in other three *E. coli* strains. There were only one strain each of *Klebsiella spp.* and *Proteus* spp. which were also found to be resistant by agar dilution method. Out of 327 gram-negative uropathogens, 298 (91.13%) strains have a MIC value under 32 µg/mL. Among 264 *E. coli* strains, 205 (77.65%) had a MIC value under MIC 16 µg/mL. All the *Enterococcus* spp. were found to have a MIC value under 32µg/mL [Table/Fig-7]. The MIC distribution of fosfomycin trometamol in *Acinetobacter* spp. were 16-64 µg/mL in present study while previous studies show higher MIC breakpoints [33]. Even though fosfomycin was seen sensitive in disc diffusion test and very low MIC breakpoint towards *Acinetobacter* spp., it remains intrinsically resistant to fosfomycin [11].

Fosfomycin MIC determination study by agar dilution method performed in Kolkata, West Bengal has reported more than 95% susceptibility among Enterobacteriaceae and Enterococcus spp., 73.33% against Pseudomonas spp. while only 50% against Acinetobacter spp. [35]. Another similar study performed by same method at Jawaharlal Institute of Postgraduate Medical Education and Research, Puducherry has published 100% Fosfomycin susceptibility in E. coli (0/217), 94.23% in Klebsiella spp. (3/52), 64% in Enterobacter spp. (9/25) and 71.88% in Pseudomonas spp. (9/32) in their reports [36]. In current study, 96.96% (287/316) of Enterobacteriaceae were reported sensitive by agar dilution method which is similar to both studies while all the Enterococcus spp. was found to be susceptible to fosfomycin in present study. One Pseudomonas spp. (14.29%) was reported resistant to fosfomycin in current study while other studies reported only 70-74% susceptibility in their studies. Overall, the resistance to fosfomycin in present study region is comparatively lower than other parts of the country [35,36].

The increasing resistance against fluoroquinolones and co-trimoxazole, drugs which are used as empirical therapy, recommends them not be used for empirical treatment. Resistance against nitrofurantoin is uncommon and is suitable for treatment of uncomplicated lower UTIs.

#### Limitation(s)

In the present study, genetic identification techniques were not implemented. Molecular screening of resistant isolates is essential to prevent the spread of plasmid-borne resistance against fosfomycin, as the mobility gene may accelerate the dissemination of fosfomycin resistance.

## CONCLUSION(S)

High susceptibility and low MIC distribution of fosfomycin trometamol suggest it along with nitrofurantoin to be used for empirical therapy armamentarium in UTIs among patient visiting day care facilities.

#### REFERENCES

- [1] Sultan A, Rizvi M, Khan F, Sami H, Shukla I, Khan HM. Increasing antimicrobial resistance among uropathogens: Is fosfomycin the answer? Urol Ann. 2015;7(1):26-30.
- [2] Pezzlo MA. Laboratory diagnosis of urinary tract infections: Guidelines, challenges, and innovations. Clinical Microbiology Newsletter. 2014;36(12):87-93.
- [3] Mehnert-Kay SA. Diagnosis and management of uncomplicated urinary tract infections. Am Fam Physician. 2005;72(3):451-56.
- Grude N, Tveten Y, Kristiansen BE. Urinary tract infections in Norway: Bacterial [4] etiology and susceptibility. A retrospective study of clinical isolates. Clin Microbiol Infect. 2001;7(10):543-47.
- Hoban DJ, Nicolle LE, Hawser S, Bouchillon S, Badal R. Antimicrobial susceptibility [5] of global inpatient urinary tract isolates of Escherichia coli: Results from the Study for Monitoring Antimicrobial Resistance Trends (SMART) program: 2009-2010. Diagn Microbiol Infect Dis. 2011;70(4):507-11.
- Cassir N, Rolain JM, Brouqui P. A new strategy to fight antimicrobial resistance: [6] The revival of old antibiotics. Front Microbiol. 2014;20(5):551.
- Rosso-Fernandez C, Sojo-Dorado J, Barriga A, Lavín-Alconero L, Palacios Z, Lopez-[7] Hernandez I, et al. Fosfomycin versus meropenem in bacteraemic urinary tract infections caused by extended-spectrum beta-lactamase-producing Escherichia coli (FOREST): Study protocol for an investigator-driven randomised controlled trial. BMJ Open. 2015;5(3):e007363.
- [8] Matthews PC, Barrett LK, Warren S, Stoesser N, Snelling M, Scarborough M, et al. Oral fosfomycin for treatment of urinary tract infection: A retrospective cohort study. BMC Infect Dis. 2016;16(1):556.
- [9] Fosfomycin for UTIs. Drug Ther Bull. 2016;54(10):114-17.
- [10] Okazaki M, Suzuki K, Asano N, Araki K, Shukuya N, Egami T, et al. Effectiveness of fosfomycin combined with other antimicrobial agents against multidrugresistant Pseudomonas aeruginosa isolates using the efficacy time index assay. J Infect Chemother. 2002;8(1):37-42.
- [11] Wayne PA. Performance standards for antimicrobial susceptibility testing. Clinical and Laboratory Standard Institute (CLSI). M100-S26.USA.2016.
- [12] Chen YH, Ko WC, Hsueh PR. Emerging resistance problems and future perspectives in pharmacotherapy for complicated urinary tract infections. Expert Opin Pharmacother. 2013;14(5):587-96.
- [13] Lee DS, Lee SJ, Choe HS. Community-acquired urinary tract infection by Escherichia coli in the era of antibiotic resistance. Biomed Res Int. 2018:2018:7656752.
- Kucheria R, Dasgupta P, Sacks SH, Khan MS, Sheerin NS. Urinary tract infections: New insights into a common problem. Postgrad Med J. 2005;81(952):83-86.
- [15] Banerjee T, Anupurba S. Risk factors associated with fluoroquinolone- Resistant enterococcal UTI in a tertiary care university hospital in north India. Ind J Med Res. 2016;144(4): 604-10.
- Manjunath GN, Prakash R, Annam V, Shetty K. Changing trends in the spectrum [16] of antimicrobial drug resistance pattern of uropathogens isolated from hospitals and community patients with urinary tract infections in Tumkur and Bangalore. Int J Biol Med Res. 2011;2(2):504-07.

- [17] Schmiemann G, Gágyor I, Pradier EH, Bleidorn J. Resistance profiles of urinary tract infections in general practice-An observational study. BMC Urology. 2012;12:33.
- [18] Shortliffe LM, McCue JD. Urinary tract infection at the age extremes: Pediatrics and geriatrics. Am J Med. 2002;113 (Suppl 1A):55S-66S.
- [19] Bitsori M. Maraki S. Raissaki M. Bakantaki A. Galanakis E. Community-acquired enterococcal urinary tract infections. Pediatr Nephrol. 2005;20(11):1583-86.
- [20] Badhan R, Singh DV, Badhan LR, Kaur A. Evaluation of bacteriological profile and antibiotic sensitivity patterns in children with urinary tract infection: A prospective study from a tertiary care center. Indian J Urol. 2016;32(1):50-56.
- [21] Huttner A, Kowalczyk A, Turjeman A, Babich T, Brossier C, Eliakim-Raz N, et al. Effect of 5-day nitrofurantoin vs single-dose fosfomycin on clinical resolution of uncomplicated lower urinary tract infection in women. JAMA. 2018:319(17):1781-89.
- [22] Sanchez GV, Master RN, Karlowsky JA, Bordon JM. In vitro antimicrobial resistance of urinary escherichia coli isolates among US outpatients from 2000 to 2010. Antimicrob Agents Chemother. 2012;56(4):2181-83.
- Gorbach SL, Bartlett JG, Blacklow NR. Infectious Diseases: Urinary Tract [23] Infection. 3rd ed. Lippincott Williams and Wilkins, Philadelphia, 2004: 861-869.
- [24] Gupta K, Hooton TM, Naber KJ, Wullt B, Colgan R, Miller LG, et al. International Clinical Practice Guidelines for the treatment of acute uncomplicated cystitis and pyelonephritis in women: A 2010 update by the infectious diseases society of America and the European Society for microbiology and infectious diseases. Clinical Infectious Diseases. 2011;52(5):e103-20.
- Ghaima KK, Khalaf ZS, Abdulhassan AA, Salman NY. Prevalence and antibiotic [25] resistance of bacteria isolated from urinary tract infections of pregnant women in Baghdad hospitals. Biomedical & Pharmacol J. 2018;11(4):1989-94.
- Butler CC, O'Brienc K, Wootton M, Pickles T, Hood K, Howe R, et al. Empiric [26] antibiotic treatment for urinary tract infection in preschool children: Susceptibilities of urine sample isolates. Family Pract. 2016;33(2):127-32.
- [27] Mandal J, Acharya NS, Buddhapriya D, Parija SC. Antibiotic resistance pattern among common bacterial uropathogens with a special reference to ciprofloxacin resistant Escherichia coli. Ind J Med Res. 2012;136(5):842-49.
- [28] Jain P. Saxena N. Spectrum of antimicrobial susceptibility pattern of pathogens isolated from patients with urinary tract infections in tertiary care hospital in Hadoti region of Rajasthan. J of Evol of Med and Dent Science. 2015;4(103):2278-4748.
- Sotto A, De Boever CM, Fabbro-Peray P, Gouby A, Sirot D, Jourdan J, et al. [29] Risk factors for antibiotic-resistant Escherichia coli isolated from hospitalized patients with urinary tract infections: A prospective study. J Clin Microbiol. 2001;39(2):438-44.
- [30] Keepers TR, Gomez M, Celeri C, Krause KM, Biek D, Critchley I. Fosfomycin and comparator activity against select enterobacteriaceae, Pseudomonas, and Enterococcus urinary tract infection isolates from the United States in 2012. Infect Dis Ther. 2017;6(2):233-43.
- [31] Jadoon SA, Ahmed A, Irshad R. Spectrum of bacterial culture and drug sensitivity vs resistance in uncomplicated urinary tract infection. J Ayubmed Coll Abbottabad. 2018;30(3):432-38.
- Barry AL, Fuchs PC. In vitro susceptibility testing procedures for fosfomycin [32] tromethamine. Antimicrob Agents Chemother. 1991;35(6):1235-38.
- [33] Lu CL, Liu CY, Huang YT, Liao CH, Teng LJ, Turnidge JD, et al. Antimicrobial susceptibilities of commonly encountered bacterial isolates to fosfomycin determined by agar dilution and disk diffusion methods. Antimicrob Agents Chemother. 2011;55(9):4295-301.
- Groote VND, Fauvart M, Kint Cl, Verstraeten N, Jans A, Cornelis P, et al. [34] Pseudomonas aeruginosa fosfomycin resistance mechanisms affect noninherited fluoroquinolone tolerance. J of Med Microbiol. 2011;60(3):329-36.
- Banerjee S, Sengupta M, Sarker TK. Fosfomycin susceptibility among multidrug-[35] resistant, extended-spectrum beta-lactamase-producing, carbapenem-resistant uropathogens. Indian J Urol. 2017;33(2):149-54.
- [36] Gopichand P, Agarwal G, Natarajan M, Mandal J, Deepanjali S, Parameswaran S, et al. In vitro effect of fosfomycin on multi-drug resistant gram-negative bacteria causing urinary tract infections. Infect Drug Resist. 2019;12:2005-13.

#### PARTICULARS OF CONTRIBUTORS:

- Tutor, Department of Microbiology, SSJ IMSR, Almora, Uttarakhand, India.
- Associate Professor, Department of Surgery, Government Medical College, Haldwani, Uttarakhand, India. 2
- З. Associate Professor, Department of Microbiology, Government Medical College, Haldwani, Uttarakhand, India.
- 4 Professor, Department of Microbiology, Government Medical College, Haldwani, Uttarakhand, India.
- 5. Assistant Professor, Department of Microbiology, Government Medical College, Haldwani, Uttarakhand, India.

# NAME, ADDRESS, E-MAIL ID OF THE CORRESPONDING AUTHOR:

Dr. Vikrant Negi, Assistant Professor, Department of Microbiology, Government Medical College, Haldwani, Uttarakhand, India. E-mail: negi.vikrant@gmail.com

## AUTHOR DECLARATION:

- Financial or Other Competing Interests: None
- Was Ethics Committee Approval obtained for this study? Yes Was informed consent obtained from the subjects involved in the study? Yes
- For any images presented appropriate consent has been obtained from the subjects. NA
- Plagiarism X-checker: Sep 04, 2020 •
- Manual Googling: Mar 18, 2021
- iThenticate Software: Apr 23, 2021 (19%)

Date of Submission: Aug 28, 2020 Date of Peer Review: Sep 28, 2020 Date of Acceptance: Apr 05, 2021 Date of Publishing: Jun 01, 2021

ETYMOLOGY: Author Origin

# PLAGIARISM CHECKING METHODS: [Jain H et al.]